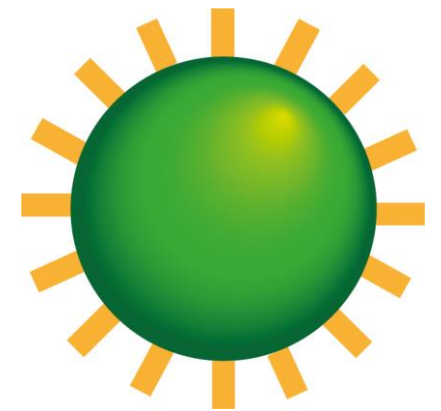


The **PANCAKE** project

Proton therapy **A**ssisted by **N**ano Pd-DDS
as **C**Ancer cell **K**illErs

Magdalena Parlinska, IFJ PAN
Olivier Seksek, IJCLab



IFJ PAN team

First name / Family name	Function (Researcher, Engineer etc)	Role in the project
Magdalena Parlinska	Senior researcher	Project leader, morphological evaluation of Pd NPs using TEM
Joanna Depciuch	Senior researcher	Synthesis of Pd NSs & NCs, functionalization NPs with anticancer drugs, Nanolive imaging
Bartosz Klębowski	Researcher	Immobilization of glucose on Pd NPs, cell & spheroid culture, proton beam irradiation

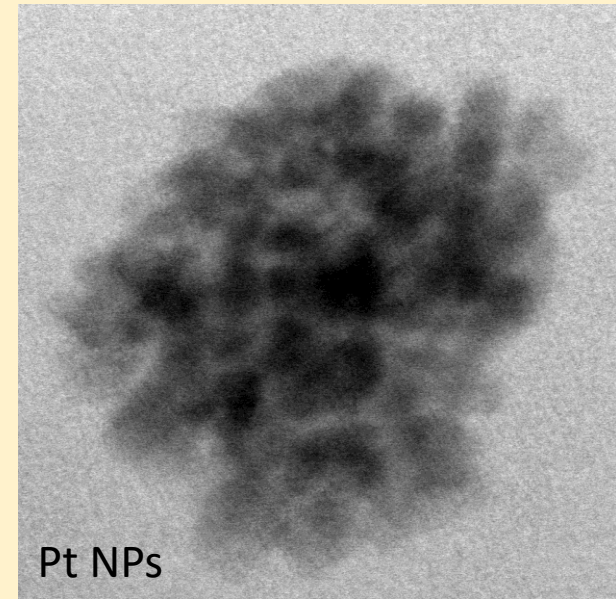
IJCLab team

First name / Family name	Function (Researcher, Engineer etc)	Role in the project
Olivier Seksek	Senior researcher	Project leader.
Joséphine Courouble	PhD student (10/23-09/26)	Data acquisition (with irradiation and NPs conditions) and data analysis.
Delphine Crépin	Science engineer	Cell lines production and characterization
Loick Ridou	Technician	Cell line production and culture (2D and 3D)
Stéphane Plaszczynski	Senior researcher	Data analysis.

IFJ PAN

Idea: The **addition of Pt NPs** to cancer cells and subsequent **proton irradiation** leads to **higher cell mortality** than **irradiation without and with Pd NPs.**

Joséphine



Experiment: In situ long-term, real-time holotomographic microscopy observations of Pd-DDS effects in cells: (i) morphological changes, (ii) absorption dynamics, (iii) interaction, (iv) accumulation sites in 3D prior and after proton irradiation.

IJCLab

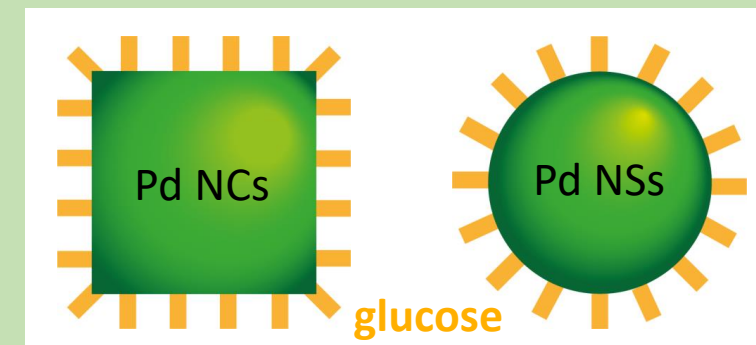
Idea: The **radioamplification by Pd-DDS** of the **X-ray irradiation (at sublethal doses)** may also lead to a **change in the individual cell behavior** inside an homogeneous or heterogeneous cell population.

Bartosz

Experiment: In situ long-term, real-time videomicroscopy data acquisition of Pd NPs, drugs and Pd-DDS effects in cells: (i) morphological changes, (ii) migration dynamics, (iii) interaction, (iv) proliferation, (v) bystander effects.

Data analysis and processing: machine learning algorithms development in order to obtain an emulator to predict cell behavior.

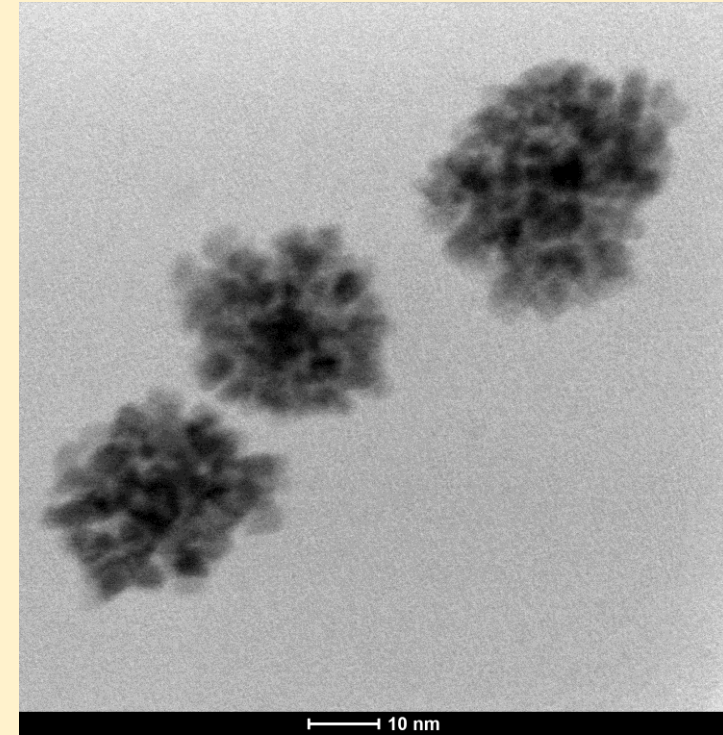
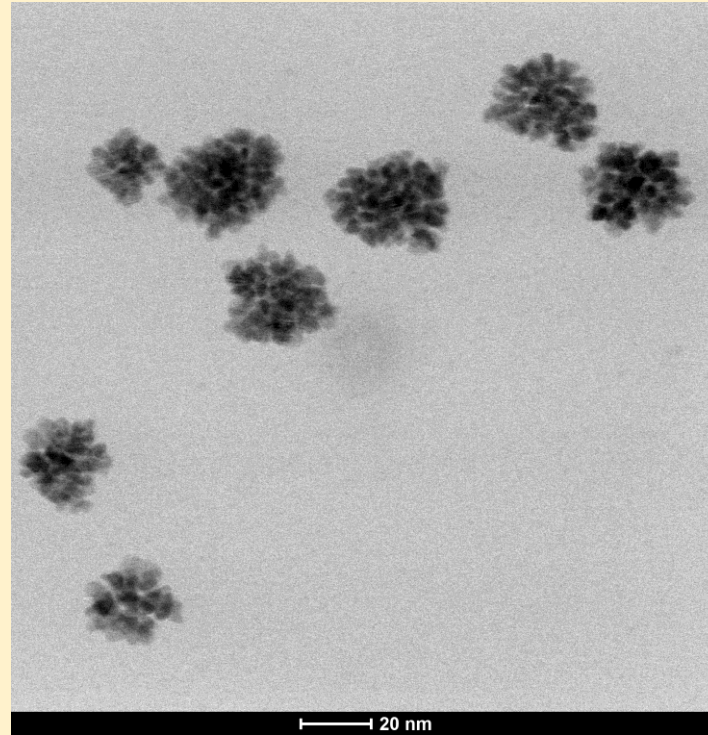
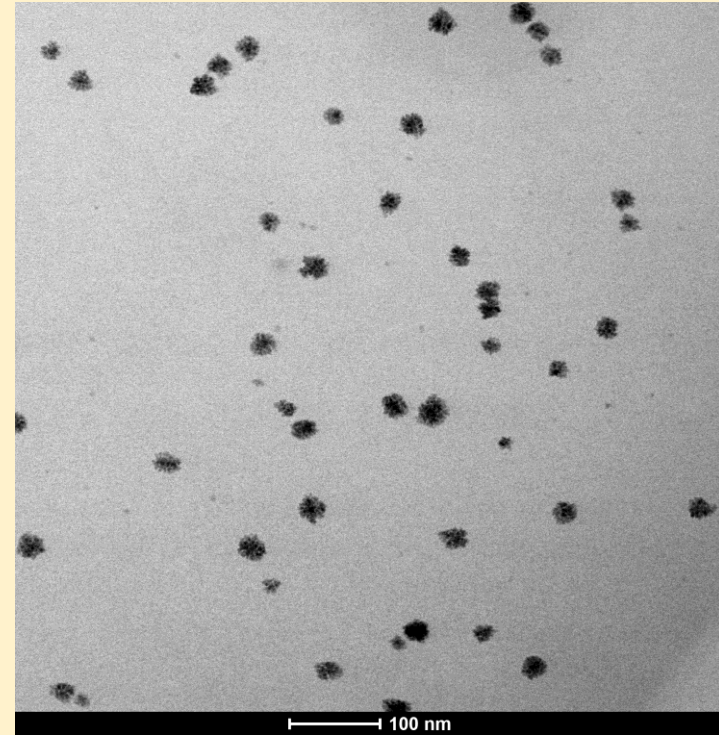
Synthesis of Pd DDS = Pd nanospheres or nanocubes functionalized by glucose immobilized on the NPs surface



**Results obtained during the visit of
Joséphine Courouble @ IFJ PAN
9. – 27.09.2024**

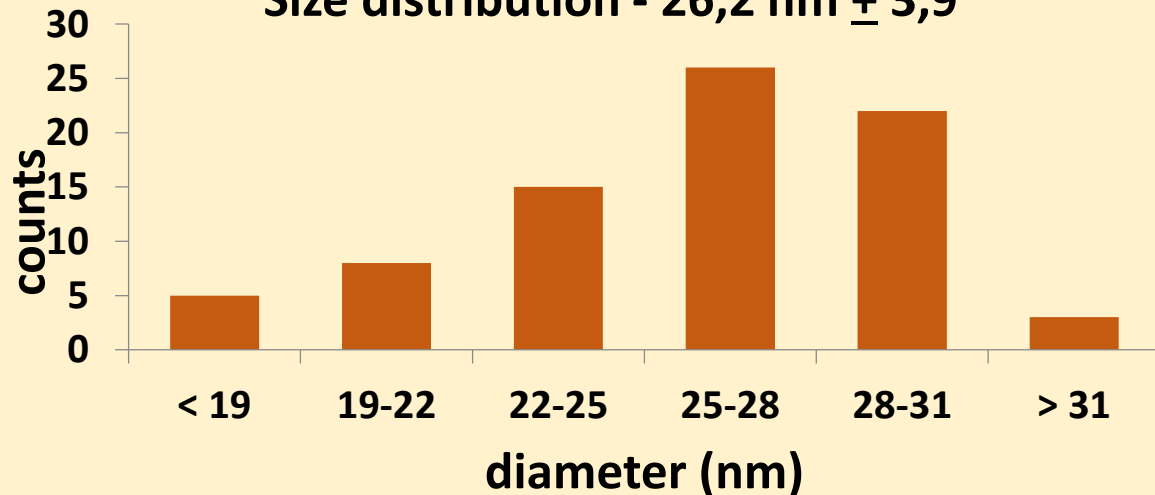
*Two months ago, Joséphine send us her cell lines, which were
cultured by Bartek to be ready for measurements*

Morphology and size distribution of Pt NPs



TEM images of the porous, flower-shaped Pt NPs

Size distribution - $26,2 \text{ nm} \pm 3,9$



2 cell lines of breast cancer MCF7 and MDA-MB 231

IJC Lab: X-ray irradiation with Pt NPs

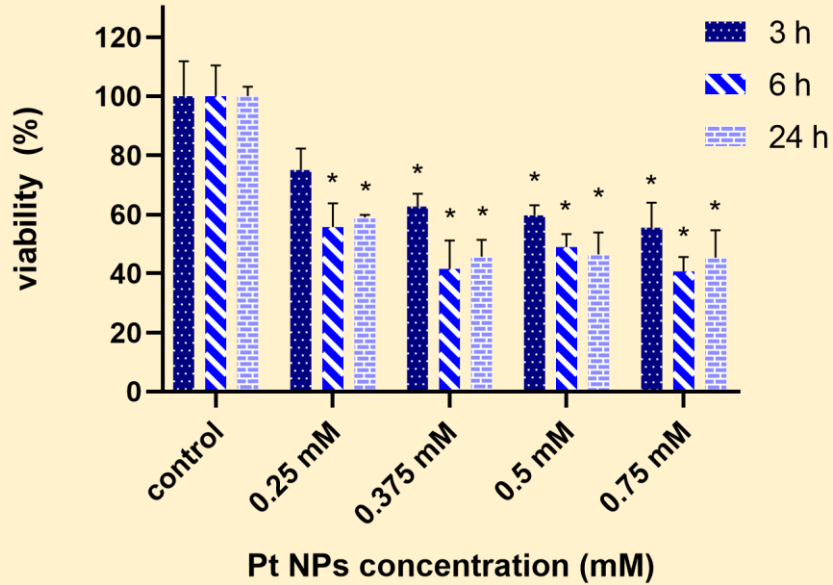
IFJ PAN: proton irradiation with Pt NPs

4 concentrations of Pt NPs:

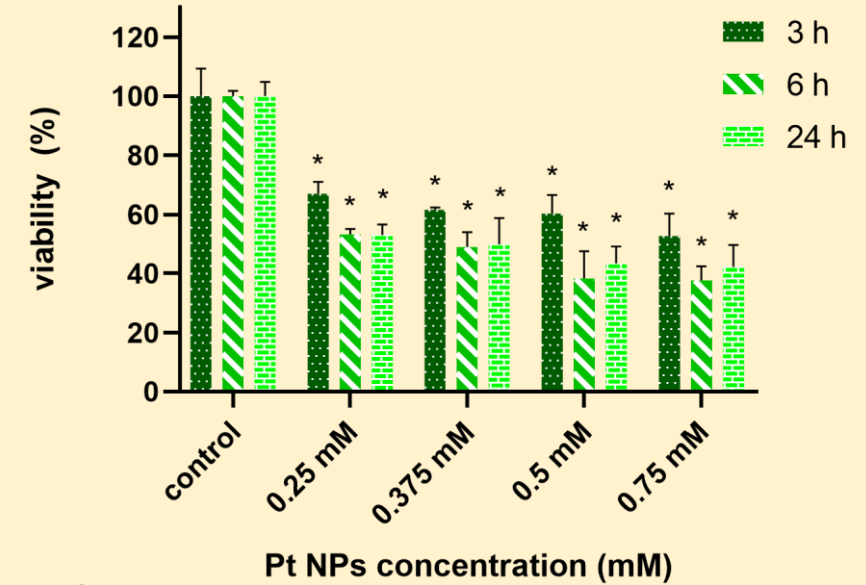
0.25 mM, 0.375 mM, 0.50 mM and 0.75 mM

MTS results – after NPs addition

MCF7 cells

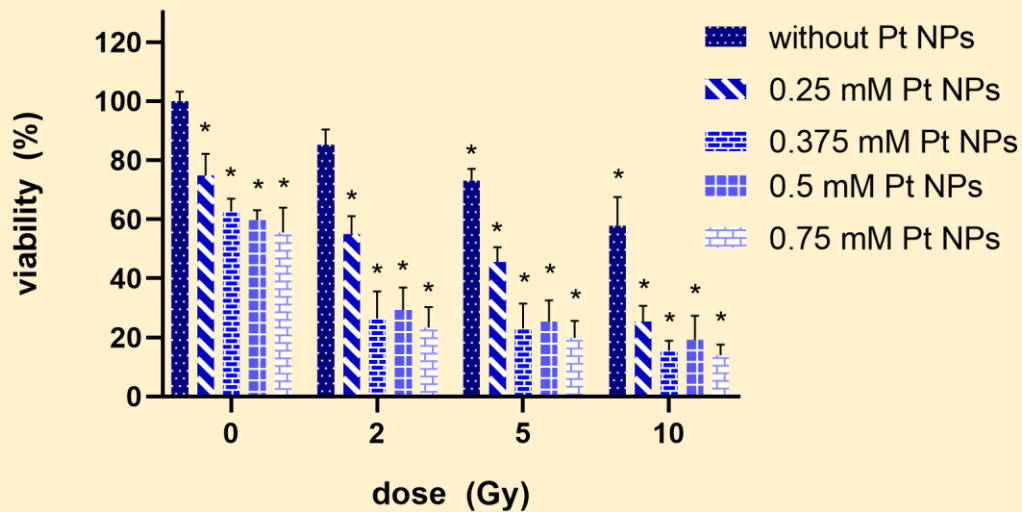


MDA MB 231 cells

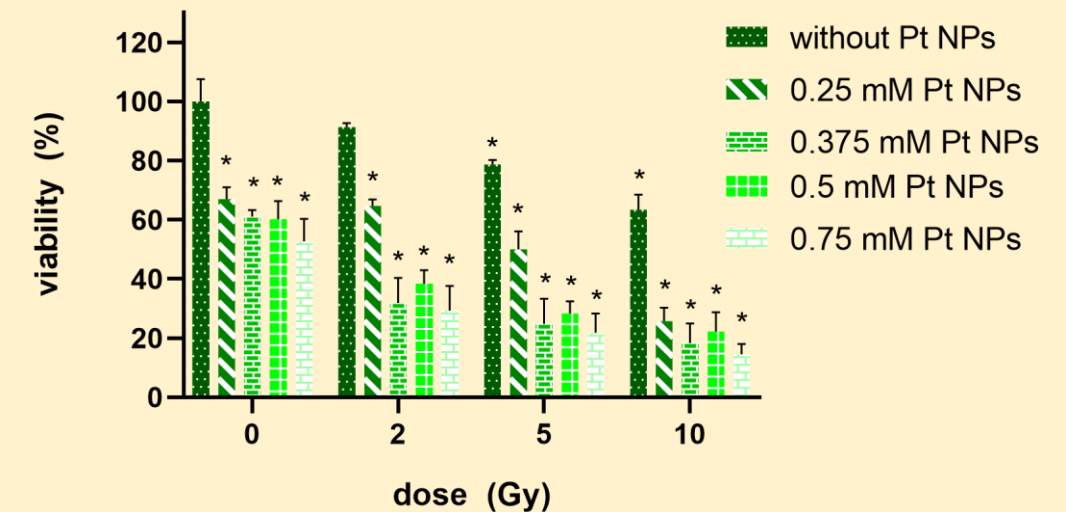


MTS results – after proton irradiation

MCF7 cells



MDA MB 231 cells



IFJ PAN

Holotomography superresolution microscope Nanolive 3d cx-a

NON-INVASIVE 3D CHARACTERIZATION

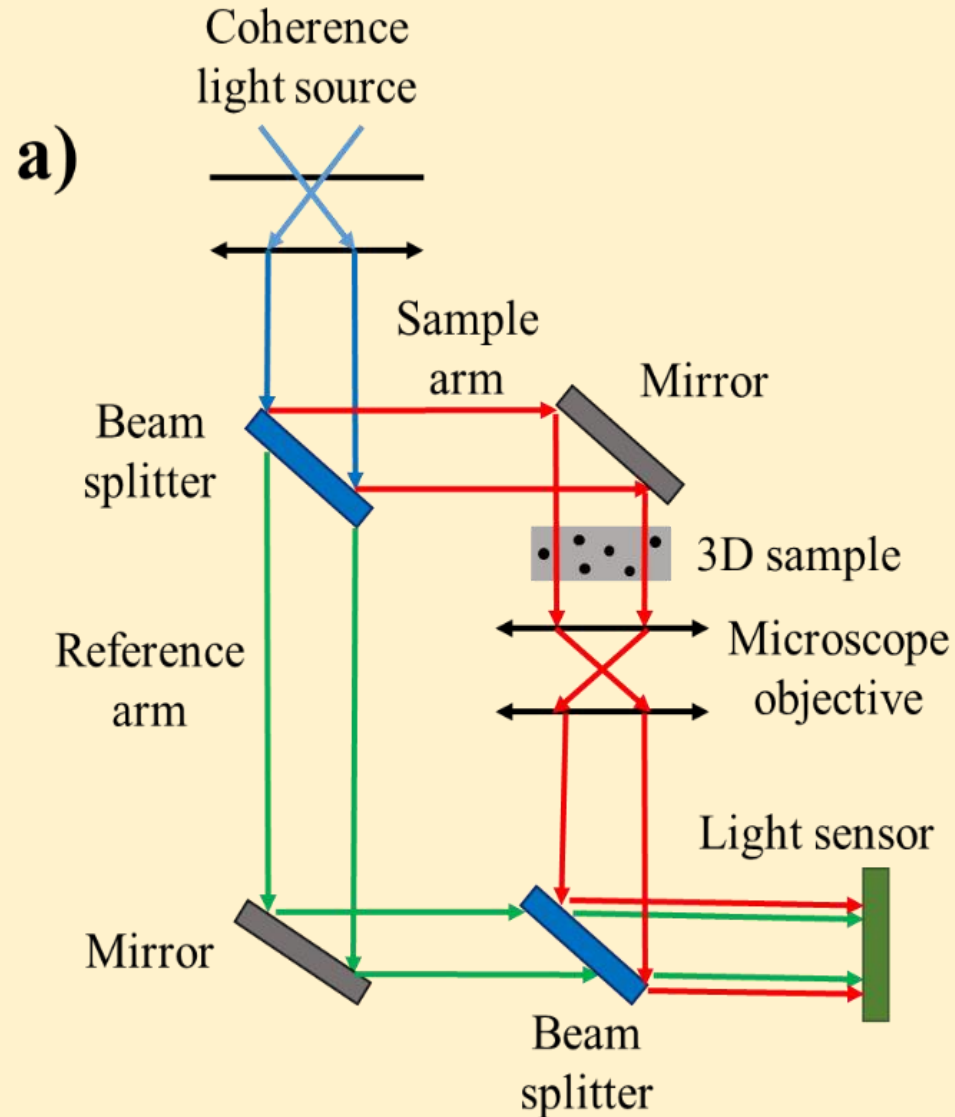
Live cell imaging in
physiological conditions
without any bleaching or
phototoxicity

LABEL-FREE 4D CONTINUOUS OBSERVATION

Measurement of cell processes
from seconds to weeks

MULTIPLEXING

High resolution and high
sensitivity characterization of
multiple cell organelles based
on their refractive index



MCF7 cells

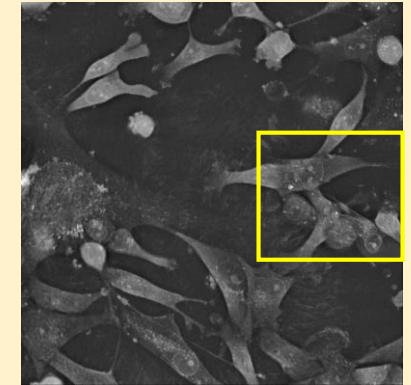
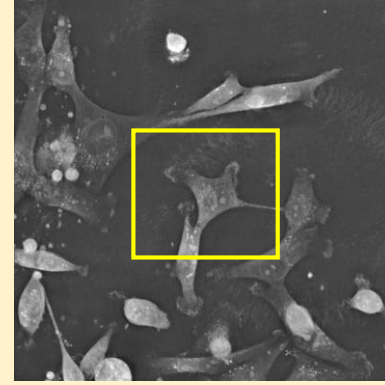
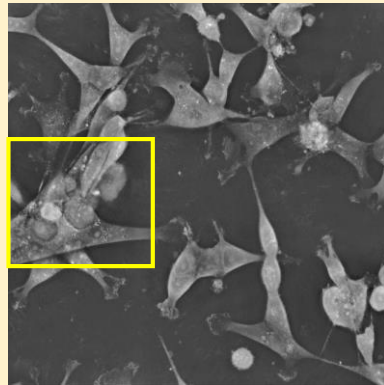
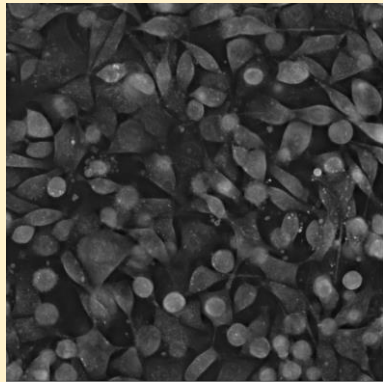
Control

0.25 mM Pt NPs

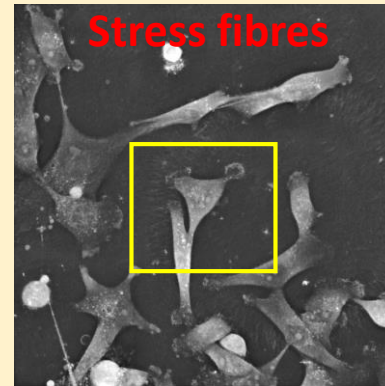
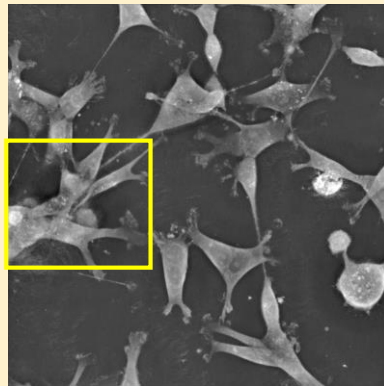
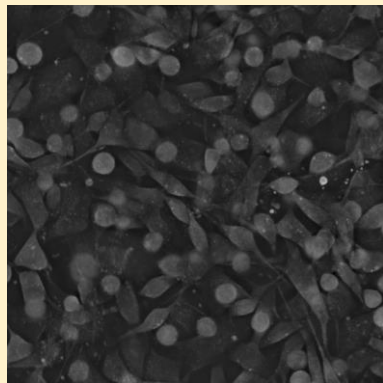
0.375 mM Pt NPs

0.5 mM Pt NPs

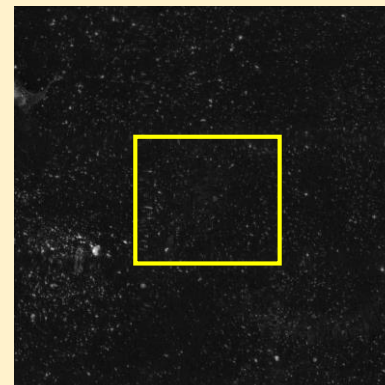
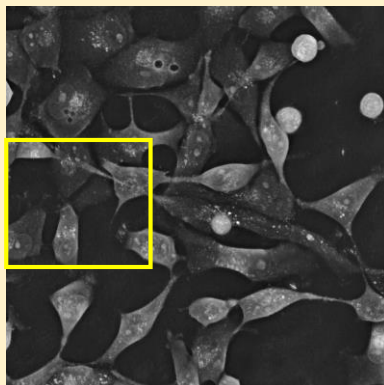
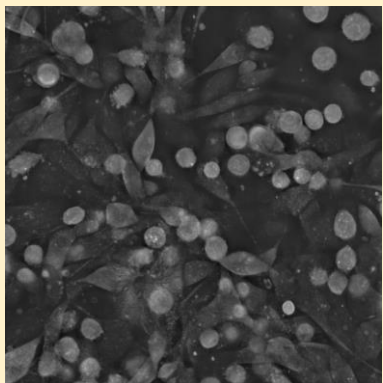
3h



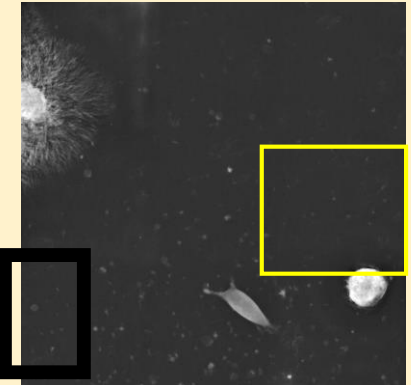
6h



24h



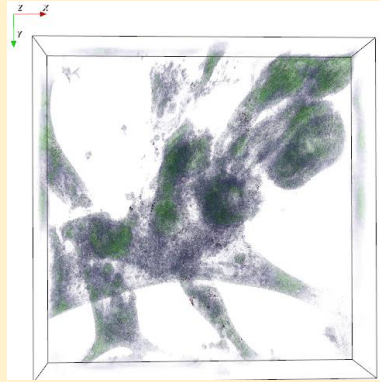
No cells visible



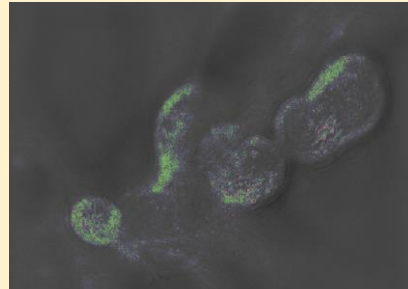
MCF7 cells + 0.25 mM Pt NPs

Pt NPs cell membrane
cytoplasm & cell nucleus

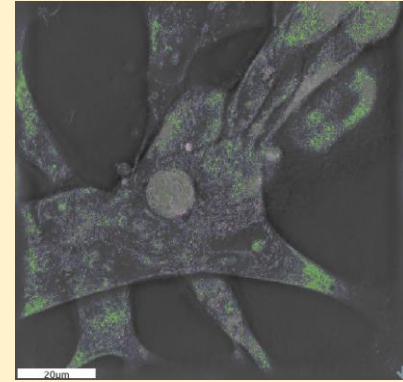
RI distribution



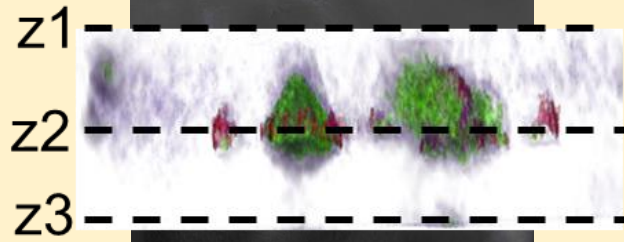
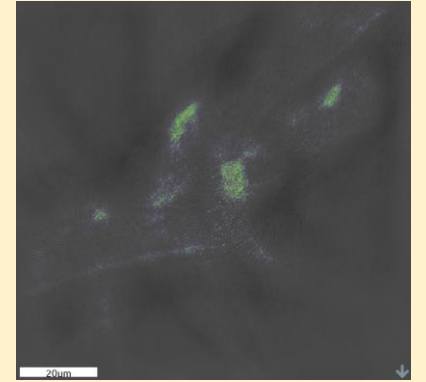
Z1



Z2



Z3



z1

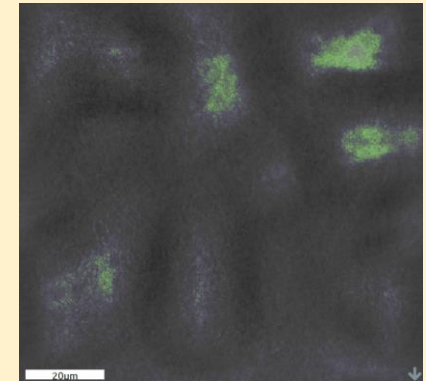
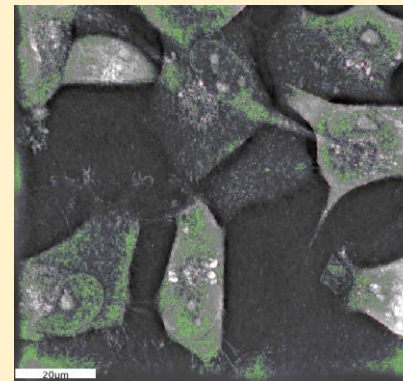
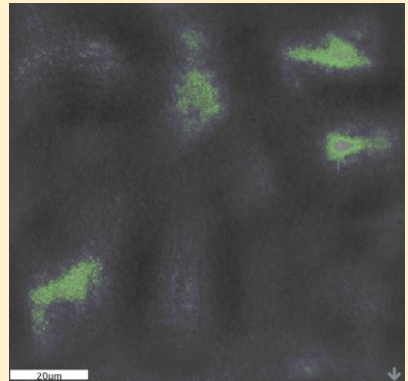
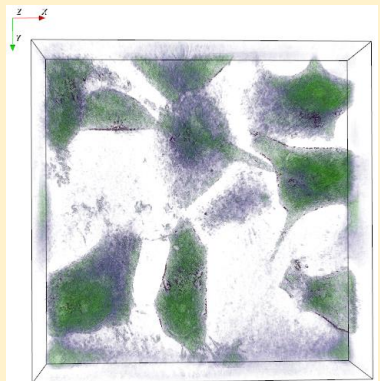
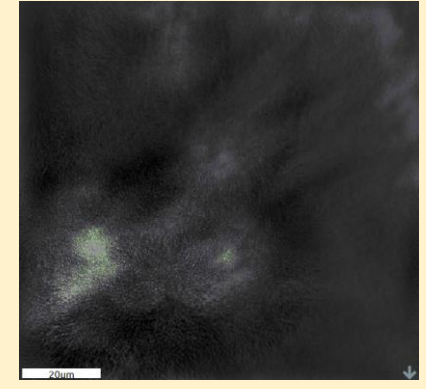
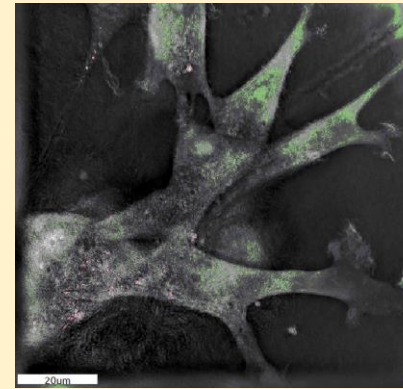
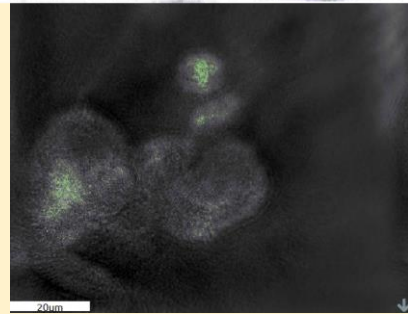
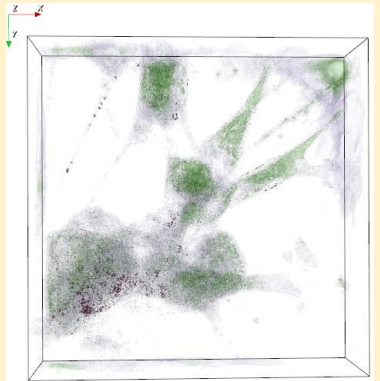
z2

z3

3h

6h

24h



MCF7 cells + 0.375 mM Pt NPs

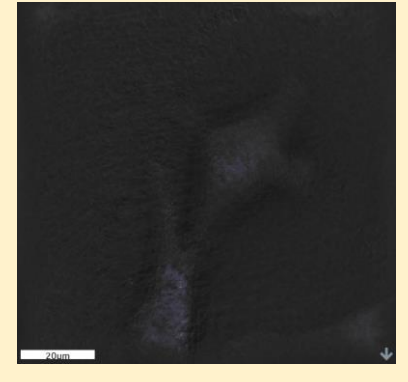
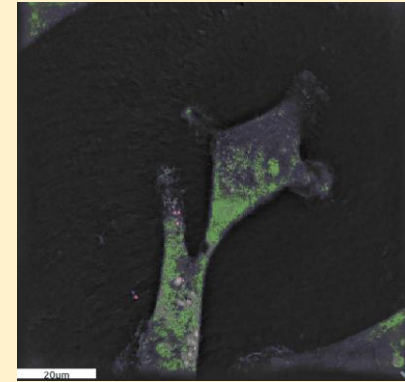
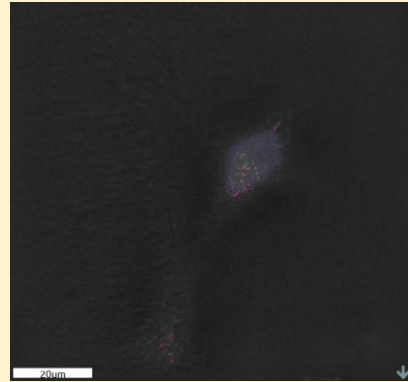
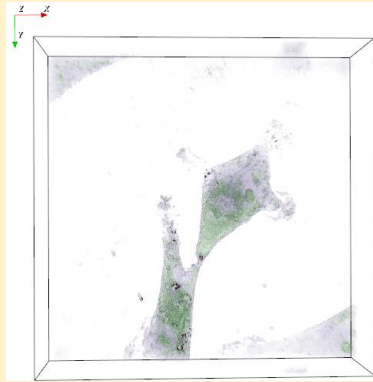
RI distribution

Z1

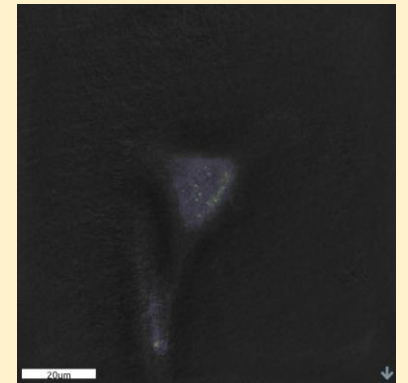
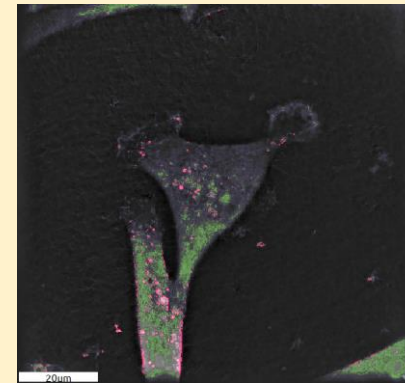
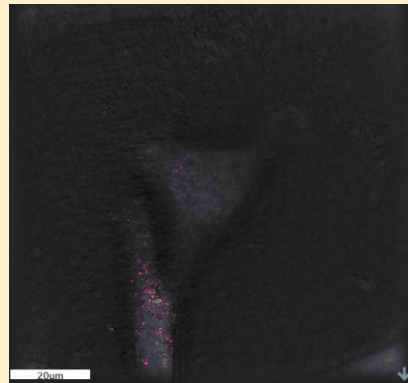
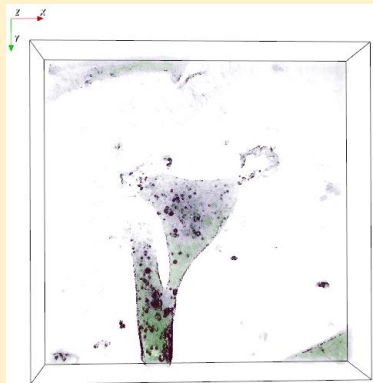
Z2

Z3

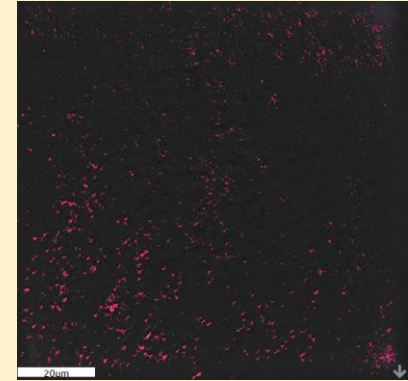
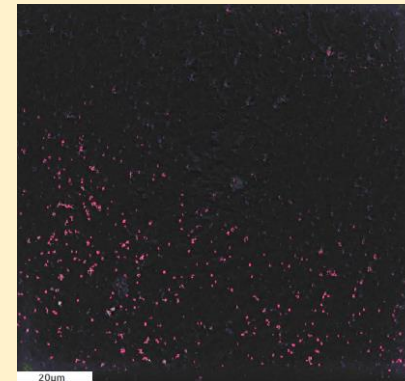
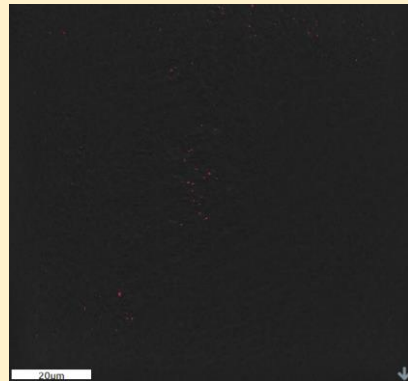
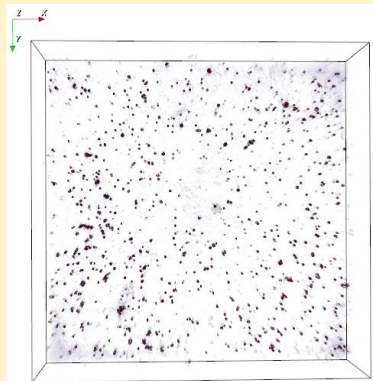
3h



6h



24h



MCF7 cells + 0.5 mM Pt NPs

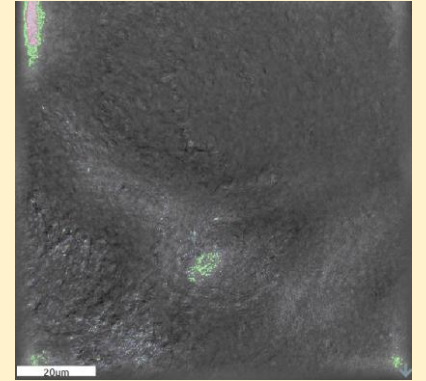
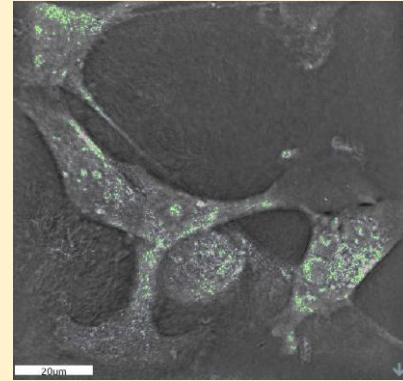
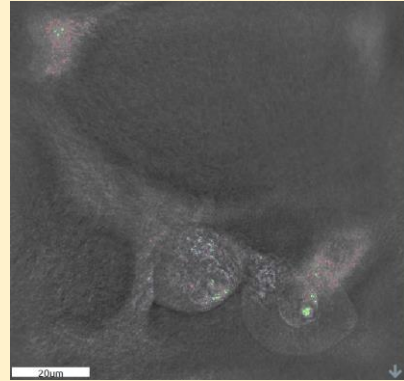
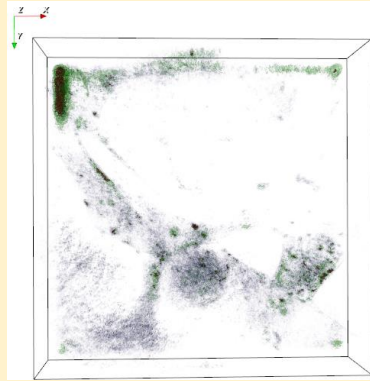
RI distribution

Z1

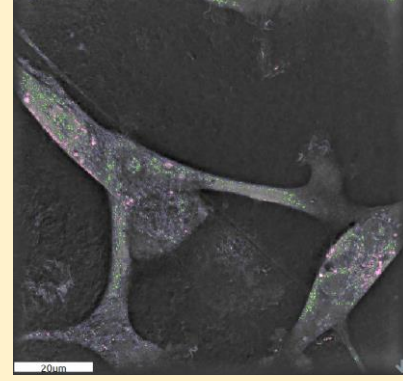
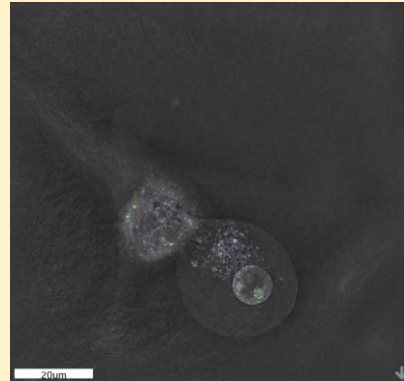
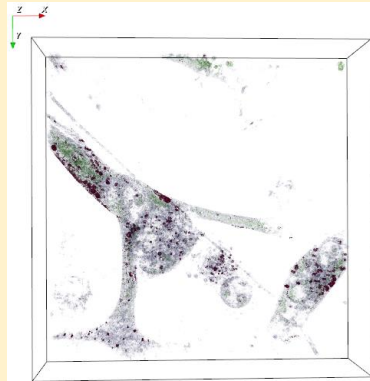
Z2

Z3

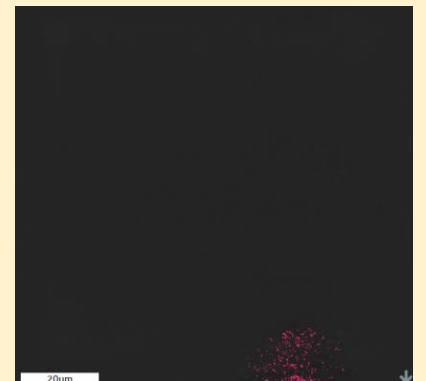
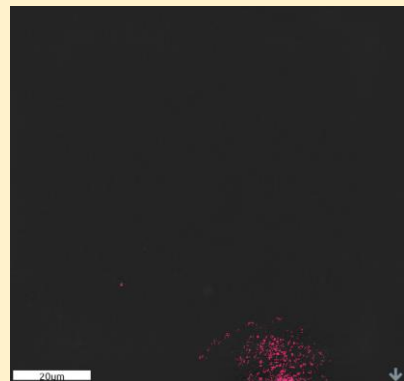
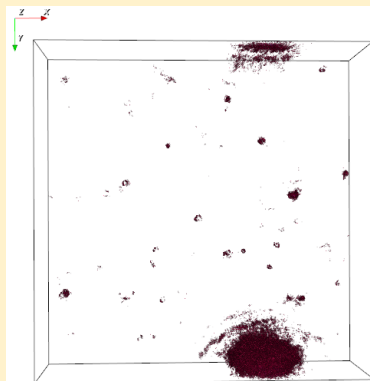
3h



6h



24h



MDA-MB 231 cells

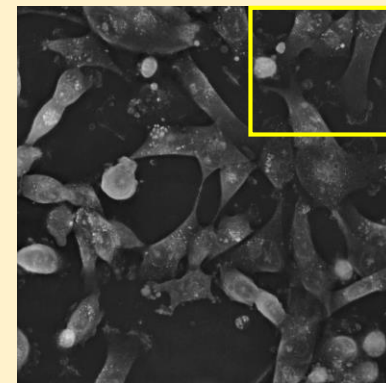
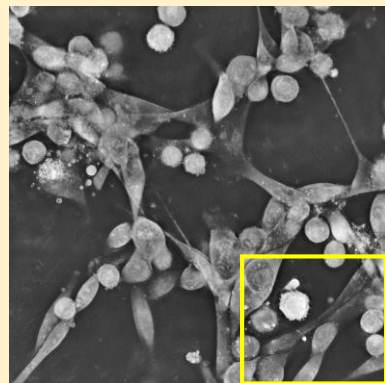
Control

0.25 mM Pt NPs

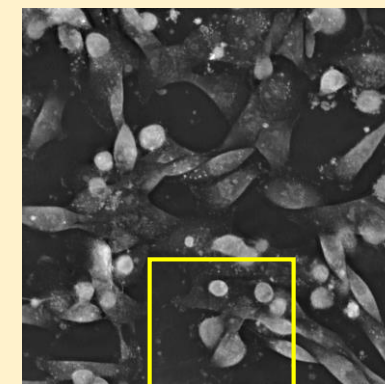
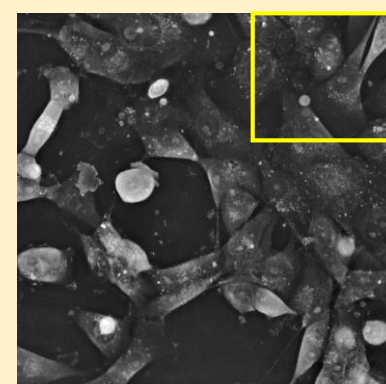
0.375 mM Pt NPs

0.5 mM Pt NPs

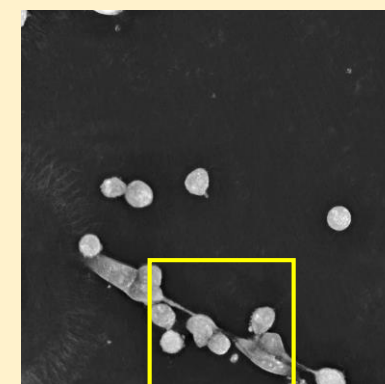
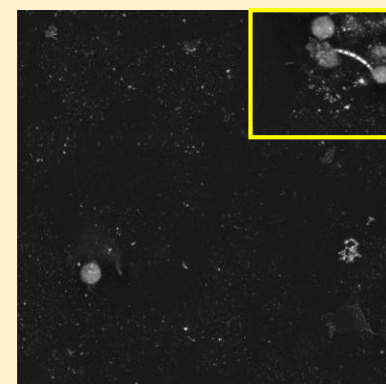
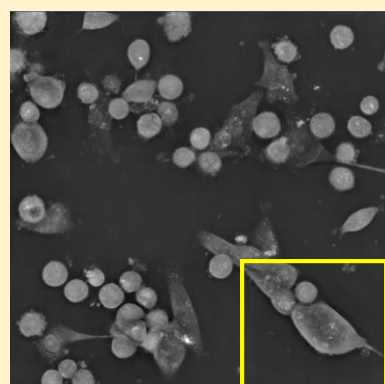
3h



6h



24h



MDA-MB 231 cells + 0.25 mM Pt NPs

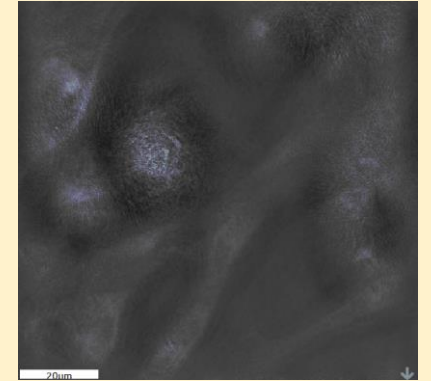
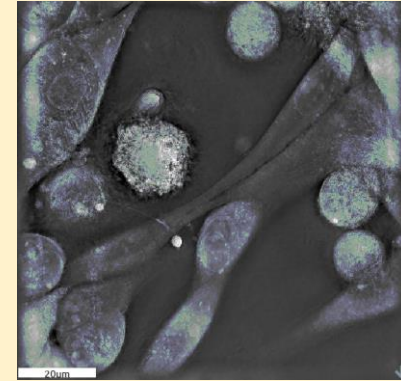
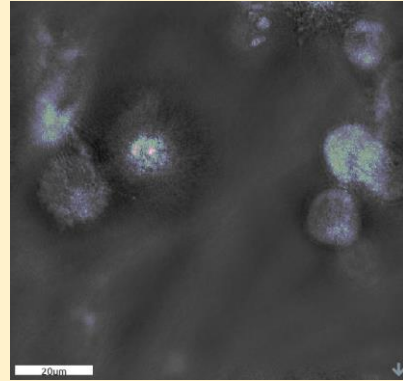
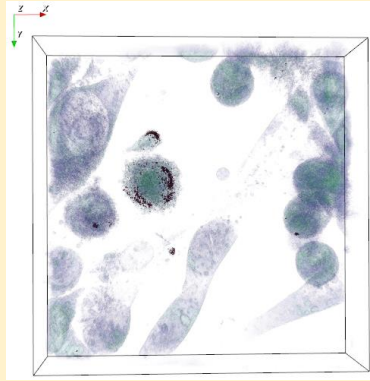
RI distribution

Z1

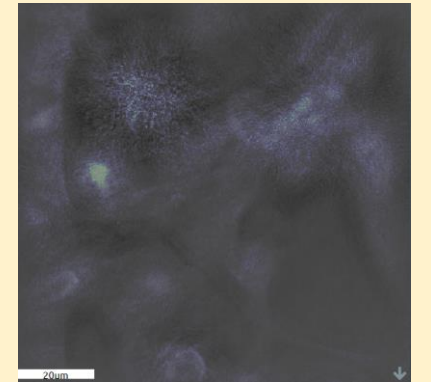
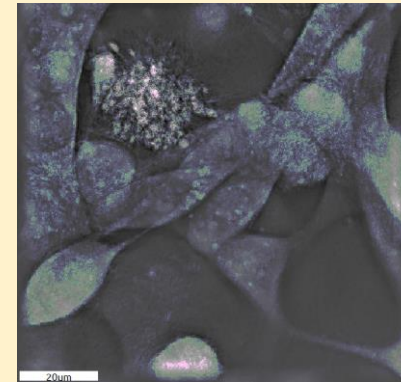
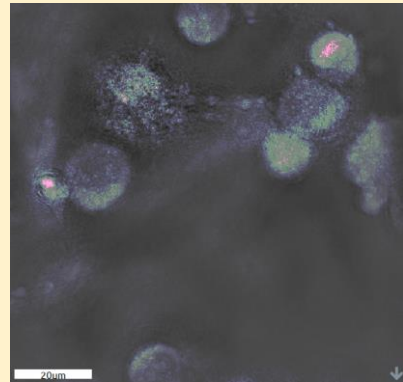
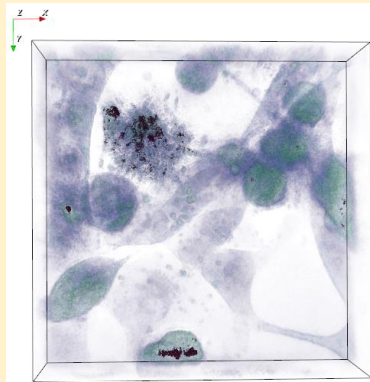
Z2

Z3

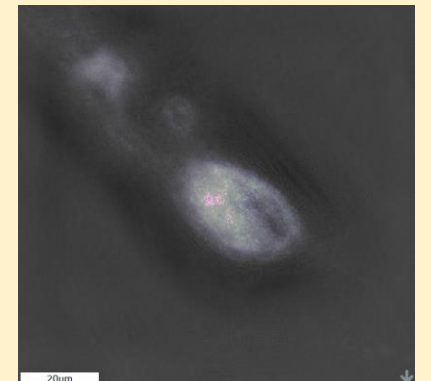
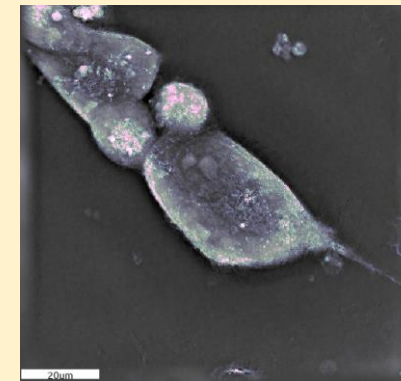
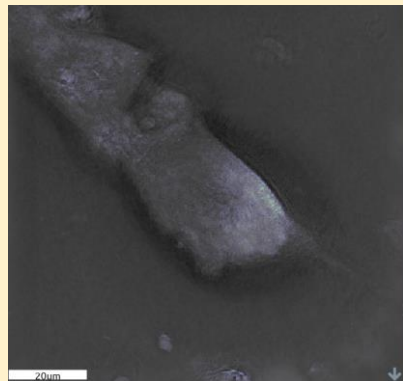
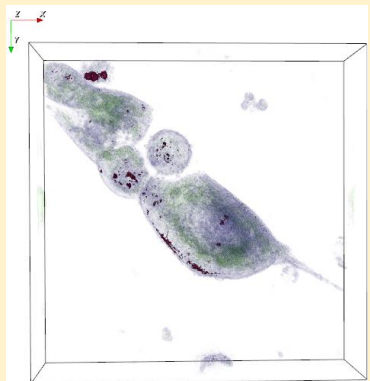
3h



6h



24h



MDA-MB 231 cells + 0.375 mM Pt NPs

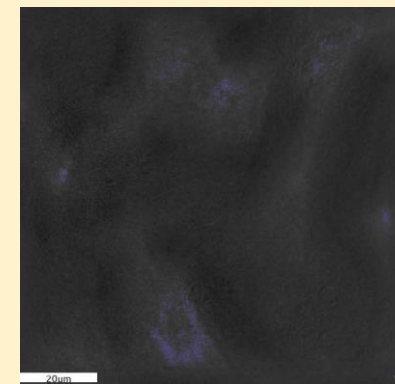
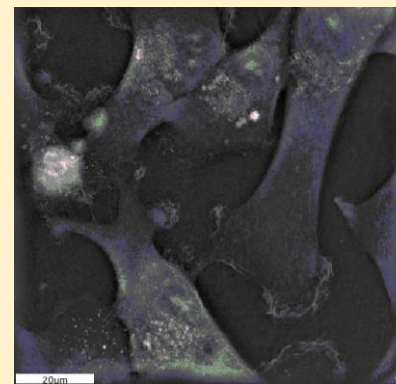
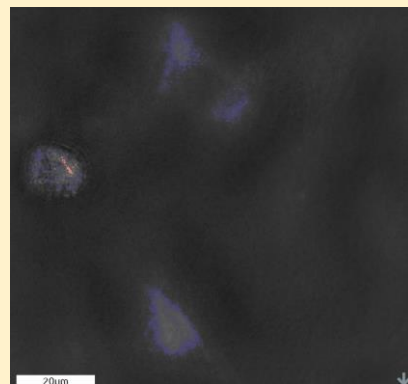
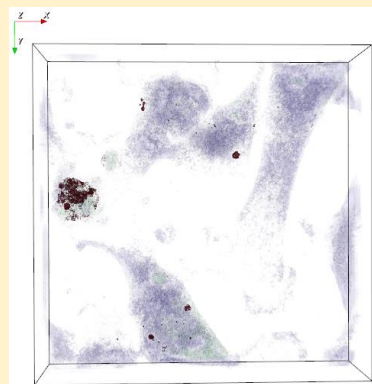
RI distribution

Z1

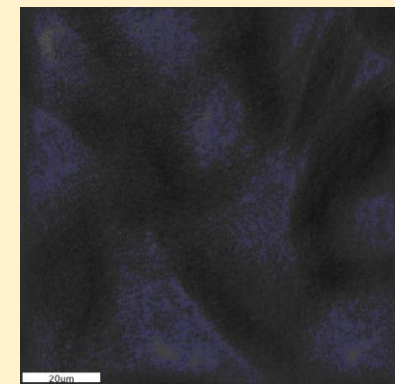
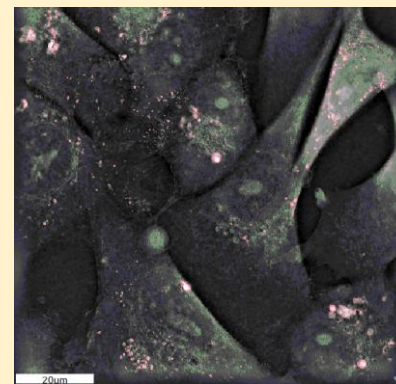
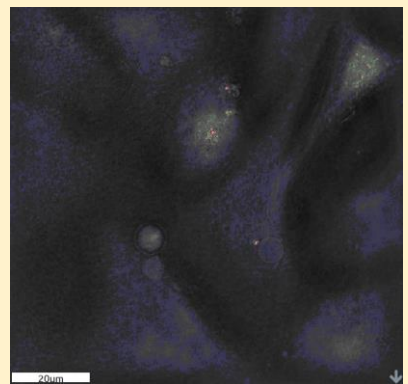
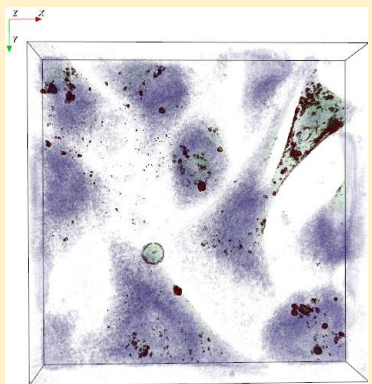
Z2

Z3

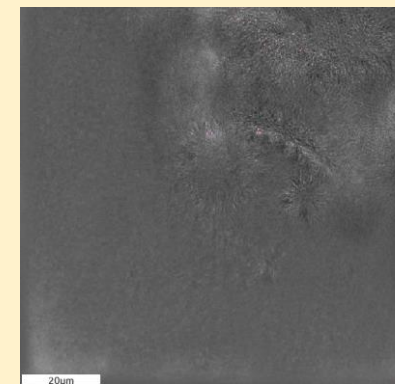
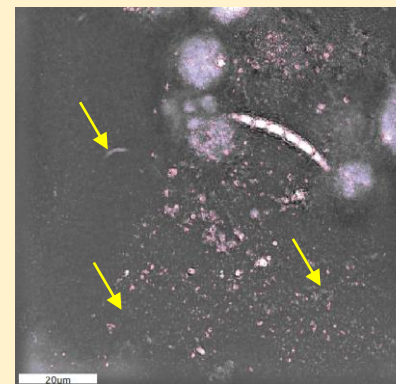
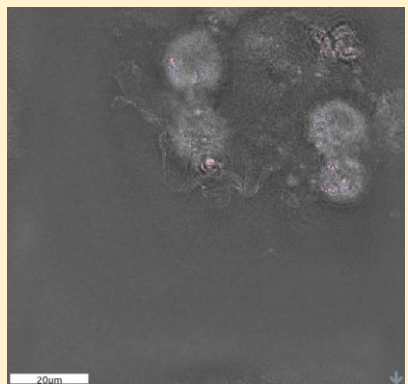
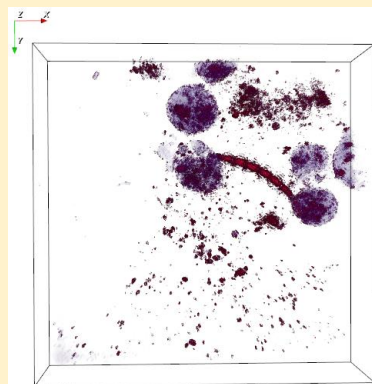
3h



6h



24h



MDA-MB 231 cells + 0.5 mM Pt NPs

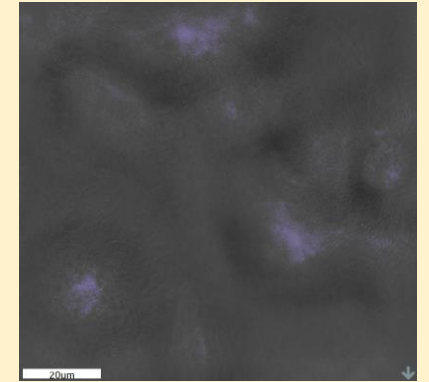
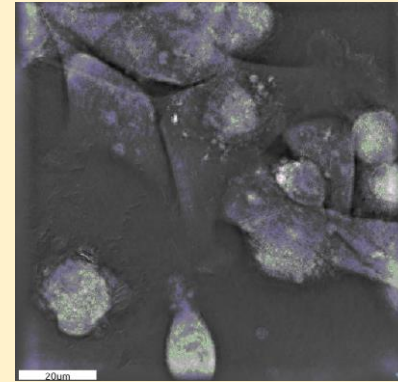
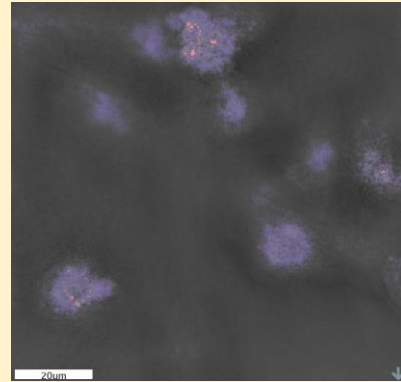
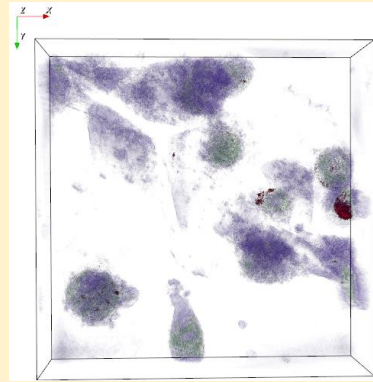
RI distribution

Z1

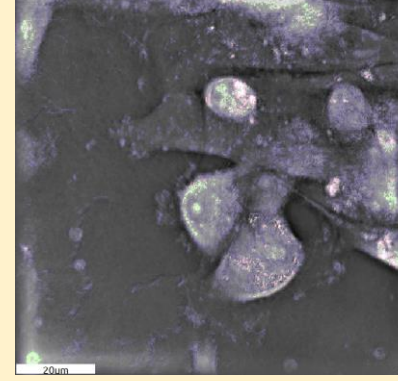
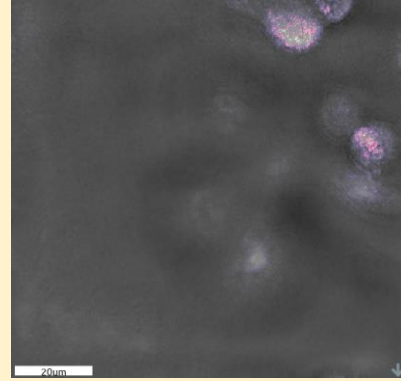
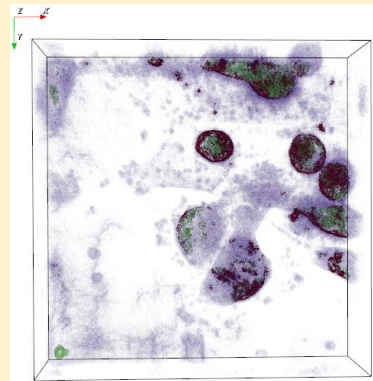
Z2

Z3

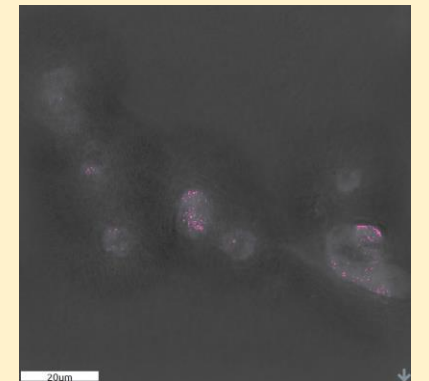
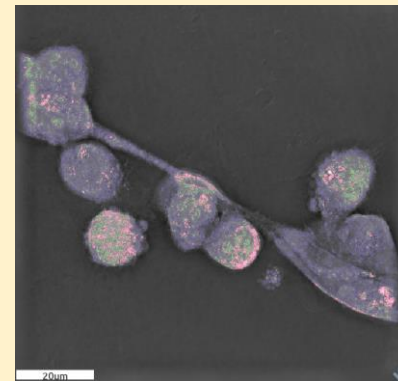
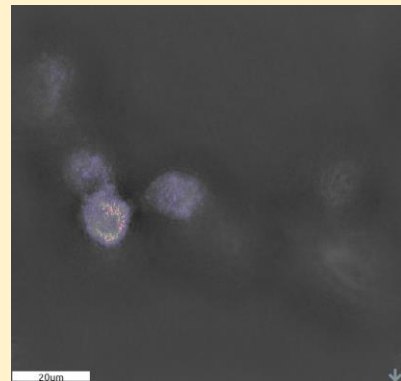
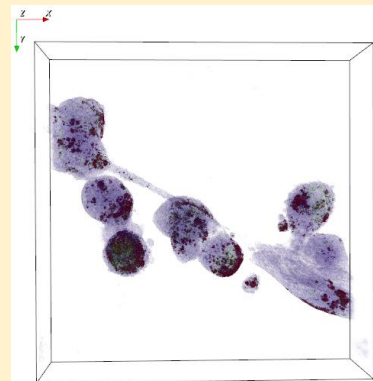
3h



6h



24h



0.25 mM Pt NPs

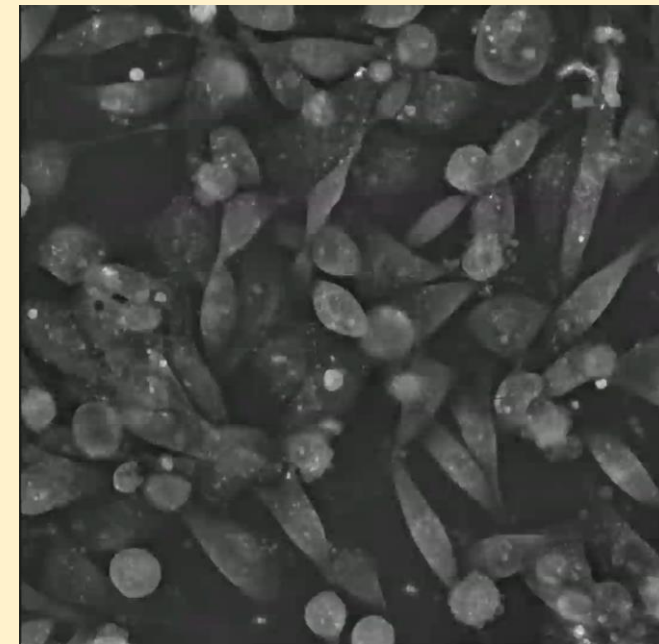
0.375 mM Pt NPs

0.5 mM Pt NPs

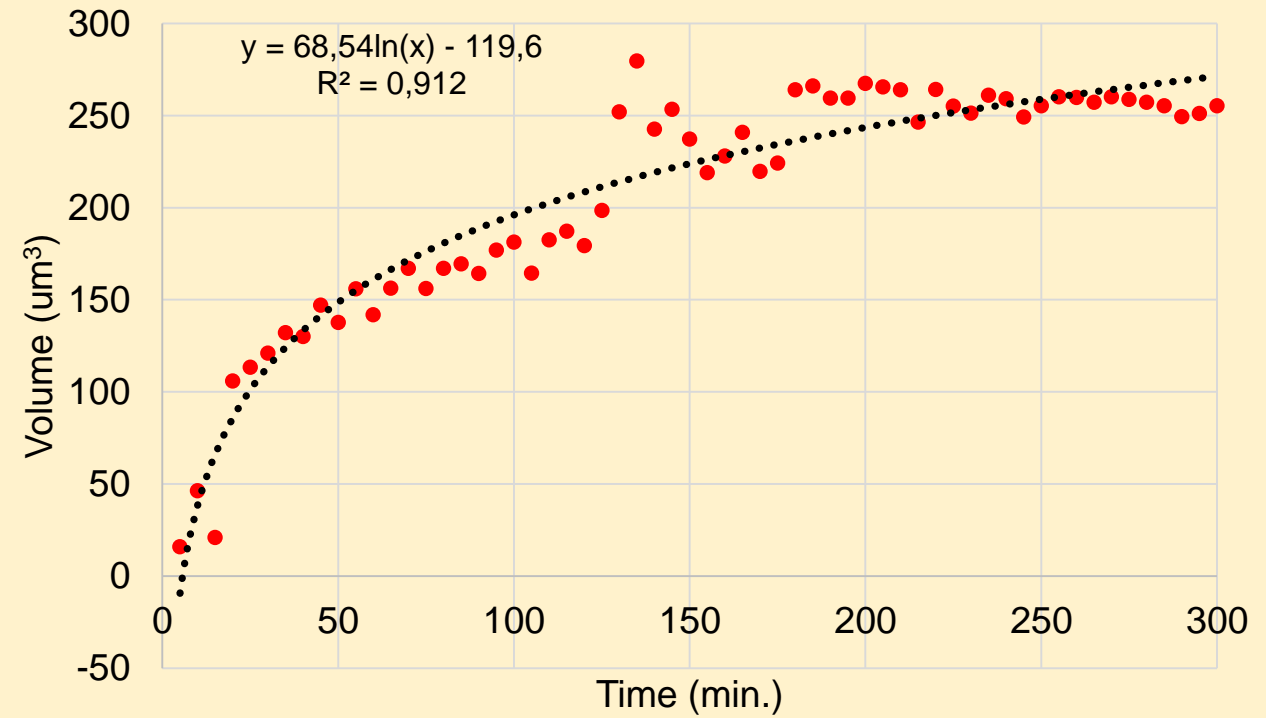
MCF7 cells



MDA-MB
231 cells



MDA-MB 231 cells + 0.25 mM Pt NPs



$$\frac{dV(t)}{dt} = S(V_{SAT} - V(t)) + D \frac{V(t)}{V_{SAT}} (V_{SAT} - V(t)), \quad (1)$$

$$V(t) = V_{SAT} \frac{1 - e^{-(S+D)t}}{1 + \frac{D}{S} e^{-(S+D)t}}, \quad (2)$$

Future plans

Dr. Bartosz Klębowski @ IJCLab 21.10 – 8.11.2024

Three months ago, Bartek send to IJCLab his cell lines, which were cultured and modified by the group of Olivier to be ready for videomicroscopy measurements

Planned measurements:

- Culture of transfected LN229 and U118 glioma cells
- X-ray irradiation of glioma cells cultured with/without spherical and cubic Pd NPs
- Analysis of control and irradiated cells using videomicroscopy

We plan to publish 2-3 publications.

Next year, we plan to submit a common project, where IJCLab (dr. Oliver Seksek) will be as a consociate.

Thank you