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## Accumulation of actinides in the mushroom *Podospora anserina*

**Pôle Énergie & Environnement – Équipe RAPHYNEE** 

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## **Actinide Behavior in the Environment**



Sources of An in the environment are various (natural and anthropogenic)



Maher, K. et al., Inorganic Chemistry 2013, 52(7), 3510-3532.

Oxidation states : one of the most important actinide mobility-governing factors

CINS

$$Pu^{4+} > Pu^{3+} > PuO_2^{2+} > PuO_2^{+}$$

strong complexes weak mobility

weak complexes strong mobility

different actinide behavior in the hydrosphere, geosphere and biosphere





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*P.* anserina: filamentous ascomycete mushroom, usually be observed on the excretion of herbivorous animals.



<u>a complete cycle (mycelial growth and sexual</u> reproduction) in the lab: 7 to 10 days at 27 ° C

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Xie et al., Fungal Genetics and Biology, 2018, 116

#### advantage

Peraza Reyes L et al. Frontiers in physiology, 2013, 4: 244.

- Simplicity of culture, short life cycle and similar physiology with macro mushrooms (extrapolation)
- Expertise in the culture and the biochemistry of *P. anserina*

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## Methodology

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Speciation in the culture medium		
Composition	Concentration (mg/L)	Concentration (M)
Mg <mark>SO</mark> 4	250	$2.08\times\mathbf{10^{-3}}$
Urea	500	$8.33\times10^{-3}$
Biotine	0.05	$2.05  imes 10^{-7}$
Thiamine	0.05	$1.89  imes 10^{-7}$
Citric acid	5	$2.60  imes 10^{-5}$
Zn <mark>SO4</mark>	5	$3.11\times10^{-5}$
Cu <mark>SO</mark> 4	0.25	$1.56\times10^{-6}$
Mn <mark>SO4</mark>	0.05	$3.31\times10^{-7}$
H <sub>3</sub> BO <sub>3</sub>	0.05	$8.06  imes 10^{-7}$
Na2M0O4·2H2O	0.05	$2.07  imes 10^{-7}$
Fe(NH <sub>4</sub> ) <sub>2</sub> ( <mark>SO<sub>4</sub></mark> ) <sub>2</sub> ·6H <sub>2</sub> O	1	$2.55  imes 10^{-6}$
Dextrin	5500	· ·

culture medium of P. anserina (pH7)

Theoretical speciation, spectroscopic techniques: XAS, UV-Vis, FTIR



\*Eu(III) as analog of Am(III) for the moment

#### **Speciation/Localization** in the mushroom

#### perithecium

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Xie et al., Fungal Genetics and Biology, 2018, 116

Microscopy / spectroscopic techniques





### **Influence of Europium concentration on the sexual reproduction**



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10 days after inoculation

#### perithecia

Open the perithecia and observe the spores

spores





few perithecia, not mature

#### No spore was observed



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#### Eu accumulation monitoring in the mycelium

• 1.1x10<sup>6</sup> estimated amount of Eu (10<sup>-5</sup>M Eu) maximum amount of Eu (10<sup>-5</sup>M Eu)  $\bigcirc$  $\begin{array}{c} \text{I} & 8.5 \times 10^{5} \\ \text{M} & 6.5 \times 10^{5} \\ \text{M} & 4.5 \times 10^{5} \\ \text{I} & 10^{5} \\ \text{M} & 10^{5} \\ \text{M} & 10^{5} \\ \text{M} & 10^{3} \\ \text{M}$ • 4.5x10<sup>5</sup> -Calculated supposing that Eu added initially was completely accumulated 5.0x10<sup>4</sup> > 3.0x10<sup>3</sup> θ  $\sim 100\%$  accumulation Calculated from the Eu remaining 1.0x10<sup>3</sup> in the medium 12 2 8 10 14 6 0 4 culture time (day)

Bioaccumulation of Eu estimated by  $[Eu]_{D0}$  -  $[Eu]_{measured}$  in the medium related to the mycelial biomass

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The maximum amount corresponds to the calculated amount when the mycelium accumulate all of the Eu present in the medium



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Imaging of Eu-loaded (5.10<sup>-4</sup>M Eu) mycelia by Fluorescence microscopy

→ Eu: homogeneously distributed in the mycelia



ATR-FTIR spectra of Eu in the culture medium (left) and Eu-loaded mycelia (right)



## **Conclusion and perspectives**





# Theoretical speciation in comparison with spectroscopic techniques



Eu accumulation monitoring in the culture medium/mycelium

#### Speciation/Localization in the mushroom

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Imaging technology in comparison with spectroscopic techniques



#### **Mechanisms of accumulation ?**

- Speciation study: characterization by spectroscopic techniques (UV-vis spectrophotometry, XAS, and TRLIFS)
- Investigation of Eu concentration in the perithecia/spore (next month)
- Adaptation of the protocol in our laboratory for further study with uranium and neptunium.

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